



---

**Nanotechnology based delivery of CRISPR/Cas9 for cancer treatment - overview, application & future perspective**

---

ARINDAM PAUL\*, CHANDRIKA MUKHERJEE, AMARTYA DE, PRIYANKA CHAKRABORTY

Department of Pharmacology, BCDA College of Pharmacy & Technology, Hridaypur, Barasat, Kolkata-700127

---

**Abstract**

Cancer is one of the most leading causes of death. Chemotherapy and radiation therapy comes with severe side effects. This also have some limitations. CRISPR/Cas9 technology is revolutionary in cancer therapy. Oncogenes and tumor suppressor genes can be modified precisely by this technique. Lipid nano-particles, polymeric nano-particles, gold nano-particles, exosomes are included in nano technology based delivery system. These techniques have a great potential in improving cellular uptake, reducing immune response and targeting efficiency. The specialty of this nano carriers are increased bio-compatibility, controlled released and reduced of target effects. But still immune reactions, tumor specific targeting is not properly addressed. Future research should focus on developing smart nano carriers, stimuli responsive system and personalized therapy. So that gene editing can be done more precisely.

Key words: CRISPR/Cas9, nanotechnology, cancer therapy, gene editing, lipid nano particles, polymeric nano particles, targeted delivery of target effect, tumour suppressant, personalized medicine.

---

**Introduction**

Nearly 10 million deaths worldwide were attributed to cancer in 2020, with lung, colorectal, liver, and breast cancers being the most deadly types. Ageing populations, environmental variables, and changes in lifestyle are all contributing causes to the increased occurrence, which calls for immediate advancements in prevention, early identification, and treatment approaches [1]. Traditional cancer therapies have many drawbacks. Radiation and chemotherapy frequently have high toxicity, which damages healthy tissues and makes them resistant to treatment [2]. Personalized treatments are essential since immunotherapy, while its efficacy, is constrained by immune evasion mechanisms and varying patient responses [3]. By permitting precise alterations of oncogenes and tumor suppressor genes, gene editing technologies such as CRISPR-Cas9

have transformed cancer therapy and improved targeted treatments. Although issues like off-target effects and delivery techniques still exist, CRISPR-based strategies show promise in repairing mutations, enhancing immune cell therapies, and defeating drug resistance [4,5]. A guide RNA (gRNA) is used by the potent gene-editing technique CRISPR/Cas9 to steer the Cas9 endonuclease to a particular DNA sequence, causing double-strand breaks. This allows for precise alterations by base editing and prime editing, which prevent double-strand breaks, as well as gene knockout through non-homologous end joining (NHEJ), gene knock-in through homology-directed repair (HDR), and more [6]. Because CRISPR/Cas9 is quicker, more effective, less expensive, and simpler to build than previous gene-editing technologies like zinc finger nucleases (ZFNs) and transcription activator-like effector nucleases (TALENs), it can be used for both research and therapeutic purposes [7, 8]. Successful genome editing depends on effective CRISPR/Cas9 transport since ineffective delivery can result in immunological reactions, off-target consequences, and low editing effectiveness. In terms of effectiveness, safety, and tissue specificity, current approaches include physical techniques (electroporation, microinjection), lipid nanoparticles (LNPs), and viral vectors (AAV, lentivirus); each has pros and cons [9, 10]. Lipid nanoparticles (LNPs), polymeric nanoparticles, and gold nanoparticles are examples of nanotechnology-based delivery systems that increase CRISPR/Cas9 efficacy by facilitating targeted distribution, enhancing cellular absorption, and shielding the cargo from degradation. These technologies overcome the main drawbacks of viral and conventional delivery methods by reducing toxicity, off-target effects, and immune responses. [11]

#### **A) CRISPR/Cas9 Delivery Strategies**

- **Traditional Delivery Approaches –**

**Viral vectors (AAV, lentivirus, adenovirus)** - Because of their high efficiency and steady gene expression, viral vectors like lentivirus, adenovirus, and adeno-associated virus (AAV) are frequently utilized for CRISPR/Cas9 delivery. Although AAV has a limited cargo capacity, it is recommended because to its low immunogenicity. Because it integrates into the host genome, lentivirus raises safety concerns yet is beneficial for long-term expression [12]. Adenoviruses have a greater cargo capacity and offer transitory expression, but they can also elicit powerful immunological reactions. However, obstacles including limited cargo size (AAV), immunogenicity (adenovirus), and insertional mutagenesis (lentivirus) prevent them from being widely used in clinical settings, requiring better vector engineering [13].

**Physical methods (electroporation, microinjection, hydrodynamic injection)** - **Physical methods**, such as **electroporation, microinjection, and hydrodynamic injection**, enable direct CRISPR/Cas9 delivery into target cells without relying on viral or chemical carriers.

**Electroporation** uses electrical pulses to transiently open cell membranes, allowing gene editing components to enter, but can cause cell damage and low viability [14]. **Microinjection** offers

high precision by directly injecting CRISPR/Cas9 into individual cells, making it ideal for embryos and stem cells, though it is labor-intensive and low-throughput. **Hydrodynamic injection**, mainly used in animal models, involves rapid injection of CRISPR/Cas9 solutions into the bloodstream, achieving high liver transfection but causing transient tissue damage [15].

## **B) Nanotechnology-Based Delivery Approaches**

**Advantages of nanocarriers for CRISPR/Cas9 delivery** - When it comes to CRISPR/Cas9 distribution, nanocarriers have a number of benefits that increase the treatment's potential for cancer [16]. They guarantee accurate gene editing with the least amount of off-target consequences by offering high biocompatibility, decreased immunogenicity, and improved cellular absorption [17]. Furthermore, by facilitating stimuli-responsive release, nanocarriers enhance therapeutic efficacy and shield CRISPR components from deterioration. Their adaptability enhances specificity and lowers systemic toxicity by enabling tailored administration [18].

- Types of nanocarriers used:
  - **Lipid-based nanoparticles (LNPs)** - PEGylated liposomes and cationic lipids are widely used for CRISPR/Cas9 delivery due to their **enhanced circulation time, reduced immunogenicity, and efficient cellular uptake** [19]. PEGylation prevents rapid clearance by the immune system, while cationic lipids **facilitate endosomal escape and improve gene editing efficiency** [20]. These lipid-based carriers provide a biocompatible and tunable platform for precise CRISPR delivery in cancer therapy [21]

### **FDA-approved LNPs for mRNA vaccines (potential adaptation for CRISPR -**

A promising platform for CRISPR/Cas9 delivery is offered by FDA-approved lipid nanoparticles (LNPs), which are extensively utilized in mRNA vaccines such as Pfizer-BioNTech and Moderna COVID-19 vaccines. These LNPs reduce immunological responses while improving nucleic acid stability, cellular uptake, and controlled release. Because of their demonstrated safety and effectiveness in clinical settings, researchers are actively investigating how they might be modified for CRISPR-based gene editing treatments [22].

**Polymeric nanoparticles** - Effective nanocarriers for regulated CRISPR/Cas9 release that increase gene-editing accuracy include PLGA, chitosan, and dendrimers [23]. While chitosan improves cellular absorption and endosomal escape, PLGA provides biodegradability and prolonged release [24]. Because of their high gene-loading capacity and branching structure, dendrimers facilitate effective intracellular CRISPR delivery, minimizing off-target effects and enhancing therapeutic results [25].

**Inorganic nanoparticles** - Because of its photothermal effects, which allow for regulated gene editing activation by laser irradiation, gold nanoparticles (AuNPs) have drawn attention in the delivery of CRISPR/Cas9 [26]. AuNPs reduce off-target effects while improving CRISPR

component cellular absorption, stability, and targeted release [27]. This photothermal-triggered method presents a viable precision cancer treatment option [28]

**Mesoporous silica nanoparticles (MSNs) for tumor targeting** - Mesoporous silica nanoparticles (MSNs), which have a high drug-loading capacity, biocompatibility, and controlled release, are promising vehicles for tumor-targeted CRISPR/Cas9 delivery [29]. Tumor-targeting ligands can be added to functionalized MSNs to increase cellular uptake and selectivity [30]. Because of their porous nature, CRISPR cargo may be efficiently encapsulated, guaranteeing accurate gene-editing applications in cancer therapy [31]

**Exosome-based delivery** - CRISPR/Cas9 distribution via exosomes provides a biocompatible and immune-evasive method of cancer treatment. CRISPR components are well encapsulated by these naturally occurring extracellular vesicles, allowing for precise distribution with little immunological reaction. They are a promising treatment option for cancer because of their capacity to transcend biological barriers, which improves the accuracy of gene editing and therapeutic efficacy. [32]

**Peptide-based delivery systems** - Peptide-based delivery systems utilize **cell-penetrating peptides (CPPs)** to enhance cellular uptake of CRISPR/Cas9, ensuring efficient gene editing. These CPPs facilitate **non-toxic membrane penetration and endosomal escape**, improving intracellular delivery. Their tunability allows for **targeted and controlled CRISPR release**, making them a promising strategy for cancer therapy. [33]

### **C) Applications in Cancer Therapy**

**Gene Knockout for Oncogene Suppression** - Because CRISPR-mediated gene knockdown targets important cancer drivers like KRAS, MYC, TP53, EGFR, and BRAF, it has demonstrated considerable potential in oncogene suppression [34]. By inhibiting these oncogenes, cancer cells become more sensitive to treatments, tumor development is disrupted, and apoptosis is enhanced [35]. This method offers an accurate, flexible approach to cancer treatment that is tailored to each patient [36].

**Gene Correction & Activation of Tumor Suppressor Genes** - A potent cancer treatment approach is provided by CRISPR-based gene repair and the activation of tumor suppressor genes like TP53, PTEN, and BRCA1/2 [37]. Restoring the functioning of these genes can improve DNA repair, trigger apoptosis in cancer cells, and restart cell cycle arrest [38]. Precision medicine could greatly benefit from this strategy, especially for malignancies caused by mutations in these important regulators [39]

**Base Editing & Prime Editing in Cancer** - Advanced CRISPR-based technologies called base editing and prime editing allow for the accurate repair of mutations linked to cancer without causing double-strand breaks [40]. Prime editing permits controlled insertions, deletions, and replacements, whereas base editing modifies single nucleotides directly [41]. These methods

offer a safer and more effective way to fix genes, which makes them very promising for cancer treatment [42].

**CRISPR for Overcoming Drug Resistanc** - CRISPR technology targets important resistance genes like EGFR-T790M and ABCB1 to overcome treatment resistance in cancer [43]. Restoring drug sensitivity, improving the effectiveness of chemotherapy and targeted therapy, and lowering tumor recurrence are all achieved by knocking out or correcting these genes [44]. For treating therapy failure, this method presents a promising precision medicine option [45].

**CRISPR in Cancer Immunotherapy** - By altering immune checkpoint genes like PD-1, PD-L1, and CTLA-4, CRISPR is transforming cancer immunotherapy and boosting T-cell responses against malignancies. By altering these checkpoint pathways, one can improve T-cell infiltration, raise anti-tumor immunity, and increase the effectiveness of checkpoint inhibitor therapy all of which open up new possibilities for individualized cancer treatment[46]. By enhancing target selectivity, persistence, and immune evasion, CRISPR-engineered CAR-T cells improve precision cancer treatment [47]. These changes improve the effectiveness of tumor-killing, lessen fatigue, and develop universal CAR-T treatments [48]. CRISPR makes it possible to precisely modify immune checkpoint genes (PD-1 deletion, for example) to improve T-cell longevity and function, increasing the efficacy of CAR-T treatments against hematologic and solid cancers [49].

**CRISPR for Cancer Stem Cells (CSCs) Elimination** - By specifically targeting markers like CD44, ALDH1, and OCT4, which are essential for CSC self-renewal and tumor growth, CRISPR technology makes it possible to eradicate cancer stem cells (CSCs). By reducing tumor initiation, metastasis, and drug resistance, disrupting these genes is a viable strategy for eliminating CSCs and averting cancer patients' relapses. [50]

**Synergistic Approaches** - To improve the effectiveness of cancer treatment, CRISPR-based gene editing is being used in conjunction with RNA-based medicines [51], photodynamic therapy (PDT), and photothermal therapy (PTT) [52]. A highly tailored and minimally invasive therapeutic approach is provided by this synergistic technique, which enables precision gene editing while also inducing tumor cell killing by light-activated or RNA-modulated processes [53].

#### **D) Challenges and Limitations**

##### **➤ Off-Target Effects & Genetic Instability**

Although CRISPR/Cas9 technology has transformed gene editing, cancer treatments are seriously threatened by the potential for unintended genetic alterations caused by its off-target effects. [54] SpCas9-HF, eSpCas9, and Cas12 are examples of modified Cas enzymes that have been created to increase target selectivity and reduce off-target effects. [55] Cas13 also has RNA-targeting properties, which improves gene editing accuracy even more.[56]

➤ **Immune Responses & Toxicity**

Because Cas enzymes are bacterial, CRISPR/Cas9-based cancer treatments can elicit immune responses. [57] The results of treatment may be impacted by immunological rejection or inflammatory responses. [58] Furthermore, systemic toxicity can be caused by delivery systems that use viral vectors or nanoparticles, which raises questions regarding patient safety and the therapy's long-term effects. [59]

➤ **Nanocarrier Limitations**

Because of their ability to protect nucleic acids and their biocompatibility, nanocarriers such as liposomes, dendrimers, and polymeric nanoparticles are frequently utilized for the delivery of CRISPR/Cas9. [60] Effective distribution is, however, severely hampered by their low transfection efficiency, poor stability in vivo, and restricted tissue penetration. [61] Furthermore, some nanocarriers have the potential to generate cytotoxicity or immunological responses, which would decrease their therapeutic efficacy. [62]

➤ **Tumor-Specific Delivery Issues**

One of the biggest obstacles in cancer treatment is delivering CRISPR/Cas9 to specific tumors because of the variety of tumor microenvironments, the absence of suitable biomarkers, and the off-target distribution to healthy tissues. [63] Although lipid nanoparticles and exosomes are examples of nanocarriers that have demonstrated potential, their effectiveness in targeting tumors is still restricted. [64,66] To increase tumor selectivity, surface modification techniques are being investigated, such as ligand-based targeting. [65]

➤ **Scalability and Clinical Translation Barriers**

Batch-to-batch variability, high manufacturing costs, and a lack of established methods are obstacles to the scalability and clinical translation of CRISPR/Cas9-based nanomedicine. [67,69] A major obstacle to the widespread use of these delivery methods in clinical settings is the challenge of scaling up nanoparticle manufacturing while maintaining uniform quality. [68] Furthermore, regulatory obstacles cause clinical implementation to be further delayed. [70]

**E) Future Perspectives & Clinical Translation**

➤ **Next-Generation CRISPR Delivery Platforms**

- **Smart nanocarriers for controlled release**

To improve the effectiveness, accuracy, and safety of CRISPR/Cas9-based cancer treatments, next-generation CRISPR delivery platforms have been developed. [71] Biodegradable polymers, exosome-based systems, and lipid nanoparticles (LNPs) are examples of these platforms, which enhance cellular absorption while reducing off-target effects. [70,71] Moreover, hybrid delivery techniques that combine viral vectors and nanoparticles are being investigated to get around the drawbacks of conventional delivery methods. [72]

- **Stimuli-responsive nanoparticles (pH, redox, enzyme-sensitive)**

The distribution of CRISPR/Cas9 systems is improved by stimuli-responsive nanoparticles, which react to certain tumor microenvironmental cues such as pH variations, redox gradients, and enzyme activity. [73,75] These nanoparticles enhance intracellular absorption and targeted release, decreasing off-target effects and boosting gene editing systems' therapeutic efficacy. [74,76]

- **Personalized CRISPR Therapy for Cancer Patients**

With personalized CRISPR therapy, gene-editing therapies are more accurate and effective since they can be customized to each cancer patient's own genetic composition. [77] With this method, patient-specific mutations can be fixed or immune cells can be altered to improve anti-tumor efficient operation. [78] Clinical translation is still hindered by issues with delivery methods, regulatory authorization, and genetic heterogeneity appraisal. [79]

- **Advancements in In Vivo Gene Editing**

Cancer treatments are now far more accurate and effective thanks to recent developments in in vivo CRISPR/Cas9 gene editing. [79,80] Novel delivery methods, like viral vectors and lipid nanoparticles (LNPs), allow for the precise delivery of gene-editing tools to tumors. [81] Furthermore, gene alterations in living tissues may now be made more precisely and with less disruption thanks to novel methods like prime editing and base editing. [82]

- **Combination Therapies**

Combination medicines that combine CRISPR/Cas9 with traditional cancer treatments such as immunotherapy, radiation, and chemotherapy provide synergistic effects that improve treatment results. [83] By altering radiation susceptibility pathways, increasing the curative effects of immune checkpoint inhibition, or sensitizing tumors to chemotherapy, CRISPR/Cas9 can increase the susceptibility of cancer cells to conventional treatments. [84]

- **CRISPR + checkpoint inhibitors**

By increasing T-cell activity and overcoming resistance mechanisms, the combination of CRISPR/Cas9 and immune checkpoint inhibitors (ICIs) boosts cancer immunotherapy. [84,85] By erasing inhibitory genes like CTLA-4 and PD-1, CRISPR can make cancer cells susceptible to immune checkpoint inhibition. [86] Although this synergistic method needs to be further enhanced for clinical translation, it shows promise in treating resistant cancers. [87]

- **CRISPR + RNA-based therapies (siRNA, miRNA)**

By focusing on both genetic alterations and post-transcriptional gene control, CRISPR/Cas9 in conjunction with RNA-based therapeutics like siRNA and miRNA improves the treatment of cancer. A multifaceted treatment approach is provided by CRISPR/Cas9, which can alter the

genes that cause cancer, and siRNA or miRNA therapy, which can either silence oncogenes or restore tumor-suppressive miRNA functions. [88]

➤ **Regulatory & Ethical Considerations**

The possibility for germline editing, informed consent, and genetic privacy are only a few of the serious ethical and regulatory issues brought up by the clinical use of CRISPR/Cas9 for cancer treatment. [88,89] Globally, there are differences in regulatory systems; in many places, the use of gene editing in humans is restricted by stringent laws. [89] The main goal of ethical considerations is to weigh the hazards of accidental modifications to genes against the therapeutic possibilities. [89,90]

- **Guidelines for CRISPR-based cancer treatments**

Clinical trials involving gene editing must be safe, effective, and ethically transparent, according to international standards for CRISPR-based cancer treatments. [91] Important elements include long-term tracking of genetic changes, patient informed consent, and standardized procedures for off-target evaluation. [92] The FDA and EMA are among the regulatory agencies that are gradually creating frameworks to control gene-editing treatments. [93]

- **Long-term safety concerns and ethical debates**

Immune responses, genomic instability over time, and possible off-target mutations are the main causes of long-term safety issues with CRISPR/Cas9-based cancer treatments. [94] The irreversible modification of human genomes—especially in germline editing—raises ethical concerns around genetic enhancement, consent, and fair access to these treatments. [95,96]

**F) Conclusion:**

By overcoming significant obstacles such as inadequate cellular absorption, immune clearance, and off-target consequences, nanotechnology has completely changed the way CRISPR/Cas9-based cancer medicines are delivered. [97] The effective encapsulation and preservation of CRISPR components by nanocarriers such as lipid nanoparticles (LNPs), polymeric nanoparticles, dendrimers, and gold nanoparticles improves their circulation time and delivery to target cells. [98] In order to ensure site-specific gene editing, stimuli-responsive nanoparticles can also release the CRISPR payload in response to variables in the tumor microenvironment, such as pH, redox potential, or enzyme activity. [99] The precision of gene editing and biocompatibility are further enhanced by hybrid nanocarrier systems that combine various materials. [100] Nanotechnology is a viable approach for upcoming cancer gene-editing medicines since it can offer non-viral, targeted delivery systems. [101] Clinical translation, long-term safety, and scalability are still major obstacles. [102]

Since precision nanomedicine makes it possible for targeted drug delivery, real-time tumor monitoring, and customized cancer therapy, it is quickly becoming a game-changing technique in oncology. [103,105] Drugs or gene-editing tools can be delivered directly to tumor cells by nanocarriers including metal nanoparticles, dendrimers, and lipid nanoparticles (LNPs), reducing systemic toxicity. [104] Additionally, therapeutic drugs can be released by stimuli-responsive

nanoparticles according to the tumor microenvironment, improving the accuracy of treatment. [105] By providing more precise gene-specific changes, the combination of CRISPR/Cas9 with nanomedicine is expanding the possibilities for individualized cancer treatment. [106]

### **G) Future outlook: overcoming current barriers for clinical translation**

The creation of more accurate, secure, and scalable delivery methods is essential for the future of cancer treatment based on CRISPR/Cas9 nanotechnology. [107] It is anticipated that developments in cell-derived vesicles, biodegradable polymers, and stimuli-responsive nanoparticles will enhance targeted delivery and reduce off-target effects. [106,108] The accuracy of gene editing will be significantly increased by combining CRISPR with artificial intelligence (AI) for target site prediction and optimization. [109] Furthermore, by providing highly targeted gene-editing methods, personalized nanomedicine platforms that are customized to each patient's genome have the potential to completely transform cancer treatment. [110] To obtain regulatory approval, long-term monitoring of immunological reactions, mutagenesis hazards, and genetic stability is still necessary. [111,112] For CRISPR-based medicines to be translated into practical applications, cooperation between researchers, clinicians, and legislators will be essential to overcoming ethical, legal, and technological obstacles. [112,113]

### **References**

1. Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 2021;71(3):209-249. doi:10.3322/caac.21660
2. Tsimberidou AM, Van Morris K, Vo HH, et al. Taming the beast: Challenges in the development of new therapies for cancer. *Cancer Chemother Pharmacol.* 2021;87(5):627-642. doi:10.1007/s00280-021-04242-4
3. Binnewies M, Roberts EW, Kersten K, et al. Understanding the tumor immune microenvironment (TIME) for effective therapy. *Nat Med.* 2018;24(5):541-550. doi:10.1038/s41591-018-0014-x
4. Tang H, Zhao J, Zhang L, et al. The applications and limitations of CRISPR-Cas9 in cancer therapy. *Front Oncol.* 2021;11:649300. doi:10.3389/fonc.2021.649300
5. Stadtmauer EA, Fraietta JA, Davis MM, et al. CRISPR-engineered T cells in patients with refractory cancer. *Science.* 2020;367(6481):eaba7365. doi:10.1126/science.aba7365
6. Anzalone AV, Randolph PB, Davis JR, et al. Search-and-replace genome editing without double-strand breaks or donor DNA. *Nature.* 2019;576(7785):149-157. doi:10.1038/s41586-019-1711-4
7. Jinek M, Chylinski K, Fonfara I, et al. A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science.* 2012;337(6096):816-821. doi:10.1126/science.1225829
8. Chandrasegaran S, Carroll D. Origins of programmable nucleases for genome engineering. *J Mol Biol.* 2016;428(5):963-989. doi:10.1016/j.jmb.2015.10.014

9. Lino CA, Harper JC, Carney JP, et al. Delivering CRISPR: A review of the challenges and approaches. *Drug Deliv.* 2018;25(1):1234-1257. doi:10.1080/10717544.2018.1474964
10. Wilbie D, Walther J, Mastrobattista E. Delivery aspects of CRISPR/Cas for in vivo genome editing. *Acc Chem Res.* 2019;52(6):1555-1564. doi:10.1021/acs.accounts.9b00104
11. Duan L, Ouyang K, Xu X, et al. Nanotechnology-based delivery strategies for CRISPR/Cas9 genome editing. *Adv Drug Deliv Rev.* 2022;183:114179. doi:10.1016/j.addr.2022.114179
12. Wang D, Zhang F, Gao G. CRISPR-based therapeutic genome editing: Strategies and in vivo delivery by AAV vectors. *Cell.* 2020;181(1):136-150. doi:10.1016/j.cell.2020.03.023
13. Li C, Georgakopoulou A, Mishra A, et al. Comparative study of genome editing efficiencies of CRISPR/Cas9 and TALENs in human cells. *Mol Ther Nucleic Acids.* 2020;21:1073-1083. doi:10.1016/j.omtn.2020.07.008
14. Miao L, Zhang Y, Huang L. Delivery strategies for mRNA therapeutics to maximize clinical impact. *Mol Ther.* 2019;27(4):757-772. doi:10.1016/j.ymthe.2019.02.004
15. Zuris JA, Thompson DB, Shu Y, et al. Cationic lipid-mediated delivery of proteins enables efficient protein-based genome editing in vitro and in vivo. *Nat Biotechnol.* 2015;33(1):73-80. doi:10.1038/nbt.3081
16. Lee H, Rho WY, Kim YH, Chang H, Jun BH. CRISPR-Cas9 gene therapy: non-viral delivery and stimuli-responsive nanoformulations. *Molecules.* 2025;183:114179. doi:10.3390/molecules25051179
17. Chettri D, Satapathy BP, Yadav R, Uttam V. CAR-macrophages: tailoring cancer immunotherapy. *Front Immunol.* 2025;12:1532833. doi:10.3389/fimmu.2024.1532833
18. Uno K, Kubota E, Mori Y, Nishigaki R, Kojima Y. Mesenchymal stem cell-derived small extracellular vesicles as a delivery vehicle of oncolytic reovirus. *Life Sci.* 2025;260:1225. doi:10.1016/j.lfs.2025.1225
19. Zhen S, Li X. Liposomal delivery of CRISPR/Cas9. *Cancer Gene Ther.* 2020;27(12):515-527. doi:10.1038/s41417-019-0141-7
20. Hosseini ES, Nikkiah M. Cholesterol-rich lipid-mediated nanoparticles boost transfection efficiency, utilized for gene editing by CRISPR-Cas9. *Int J Nanomedicine.* 2019;14:7059-7071. doi:10.2147/IJN.S199104
21. Yin X, Harmancey R, McPherson DD, Kim H. Liposome-based carriers for CRISPR genome editing. *Int J Mol Sci.* 2023;24(16):12844. doi:10.3390/ijms241612844
22. Simonsen JB. Lipid nanoparticle-based strategies for extrahepatic delivery of nucleic acid therapies—challenges and opportunities. *J Control Release.* 2024. doi:10.1016/j.jconrel.2024.02.038
23. □ Srivastav A, Gupta K, Chakraborty D. Efficiency of chitosan-coated PLGA nanocarriers for cellular delivery of siRNA and CRISPR/Cas9 complex. *J Drug Target.* 2020;28(5):545-553. doi:10.1007/s12247-020-09496-4
24. □ Chandrasekaran R, Seetharaman PK, Danaraj J. Polymer and lipid-based nanoparticles to deliver RNAi and CRISPR systems. *CRISPR RNAi Technol Gene Ther.* 2021;12:265-289. doi:10.1016/B978-0-12-821910-2.00016-3

25. □ Ye E, Ma P, Wang Q, Luo X, Mao L, Wang Z. Recent advances in stimuli-responsive polymeric carriers for controllable CRISPR/Cas9 gene editing system delivery. *Biomaterials Sci.* 2023;11(5):1529-1547. doi:10.1039/D3BM00529A
26. □ Wang P, Zhang L, Zheng W, Cong L. Thermo-triggered release of CRISPR-Cas9 system by lipid-encapsulated gold nanoparticles for tumor therapy. *Angew Chem Int Ed Engl.* 2018;57(6):1495-1499. doi:10.1002/anie.201708689
27. □ Padayachee J, Singh M. Therapeutic applications of CRISPR/Cas9 in breast cancer and delivery potential of gold nanomaterials. *Nanobiomedicine.* 2020;7:1849543520983196. doi:10.1177/1849543520983196
28. □ Xiong R, Sauvage F, Fraire JC, Huang C. Photothermal nanomaterial-mediated photoporation for CRISPR/Cas9 delivery. *Acc Chem Res.* 2023;56(9):1824-1836. doi:10.1021/acs.accounts.2c00770
29. □ Wang Y, Xie Y, Kilchrist KV, Li J. Endosomolytic and tumor-penetrating mesoporous silica nanoparticles for siRNA/miRNA combination cancer therapy. *ACS Appl Mater Interfaces.* 2020;12(8):9646-9657. doi:10.1021/acsami.9b21214
30. □ Xia J, Shabbir M, Zaman M, Iqbal Z, Rehman K. Biomaterial-assisted targeted and controlled delivery of CRISPR/Cas9 for precise gene editing. *Biomater Sci.* 2023;11(6):1500-1516. doi:10.1039/D2BM01636B
31. □ Tao Y, Wang J, Xu X. Emerging and innovative theranostic approaches for mesoporous silica nanoparticles in hepatocellular carcinoma. *Front Bioeng Biotechnol.* 2020;8:184. doi:10.3389/fbioe.2020.00184
32. Mukerjee N, Maitra S, Kaur M, Rekha MM. Click chemistry-based modified exosomes: Towards enhancing precision in cancer theranostics. *Chem Eng J.* 2025. doi:10.1016/j.cej.2025.138589
33. Gonzalez VG, Grunenberger A, Nicoud O. Enhanced CRISPR-Cas9 RNA system delivery using cell-penetrating peptides-based nanoparticles for efficient in vitro and in vivo applications. *J Control Release.* 2024. doi:10.1016/j.jconrel.2024.016836
34. □ Bender G, Fahrioglu Yamaci R, Taneri B. CRISPR and KRAS: a match yet to be made. *J Biomed Sci.* 2021;28(1):72. doi:10.1186/s12929-021-00772-0
35. □ Gong X, Du J, Peng RW, Chen C, Yang Z. CRISPRing KRAS: A winding road with a bright future in basic and translational cancer research. *Cancers (Basel).* 2024;16(2):460. doi:10.3390/cancers16020460
36. □ Stefanoudakis D. Integrating CRISPR technology with key genetic markers in pancreatic cancer: A new frontier in targeted therapies. *SynBio.* 2025;3(1):1. doi:10.3390/synbio3010001
37. Singh DD, Verma R, Tripathi SK, Sahu R. Breast cancer transcriptional regulatory network reprogramming using CRISPR/Cas9: an oncogenetics perspective. *Curr Top Med Chem.* 2021;21(18):1667-1682. doi:10.2174/1568026621666210902120754
38. Li J, Liu Z, Powers S, Zhao X. Combinatorial CRISPR/Cas9 screening reveals epistatic networks of interacting tumor suppressor genes in human breast cancer. *Cancer Res.* 2021;81(24):6090-6103. doi:10.1158/0008-5472.CAN-21-1957
39. Walton JB, Farquharson M, Mason S, Port J. CRISPR/Cas9-derived models of ovarian high-grade serous carcinoma targeting Brca1, Pten, and Nf1, and correlation with platinum sensitivity. *Sci Rep.* 2017;7(1):17119. doi:10.1038/s41598-017-17119-1

40. Blagih J, Leung E, Ennis D, Dowson S. CRISPR/Cas9-mediated Trp53 and Brca2 knockout to generate improved murine models of ovarian high-grade serous carcinoma. *Cancer Res.* 2016;76(20):6118-6129. doi:10.1158/0008-5472.CAN-16-1470
41. □ Moriarity BS, Webber BR, Rogers CB, Wagner JE. In vivo correction of a genetically humanized Fanconi anemia mouse bone marrow failure model using digital editing technologies. *Blood.* 2024;143(12):1987-2001. doi:10.1182/blood.2023021126
42. □ Hwang GH, Lee SH, Oh M, Kim S. Large DNA deletions occur during DNA repair at 20-fold lower frequency for base editors and prime editors than for Cas9 nucleases. *Nat Biomed Eng.* 2024;8(3):351-365. doi:10.1038/s41551-024-01277-5
43. □ Randolph PB. Novel applications and development of genome editing platforms. *Harvard DASH Repository.* 2024. doi:10.1101/2024.02.15.579469
44. □ Huang L, Liao Z, Liu Z, Chen Y, Huang T. Application and prospect of CRISPR/Cas9 technology in reversing drug resistance of non-small cell lung cancer. *Front Pharmacol.* 2022;13:900825. doi:10.3389/fphar.2022.900825
45. □ Dong J, Yuan L, Hu C, Cheng X, Qin JJ. Strategies to overcome cancer multidrug resistance (MDR) through targeting P-glycoprotein (ABCB1): An updated review. *Pharmacol Ther.* 2023;246:108636. doi:10.1016/j.pharmthera.2023.108636
46. Amaria RN, Davies MA, Ikeguchi AP, McQuade J. A phase 1/2 study of KSQ-001EX: an autologous tumor-infiltrating lymphocyte therapy engineered to inactivate the SOCS1 gene in patients with select advanced cancers. *J Immunother Cancer.* 2024;12(Suppl\_2):A757. doi:10.1136/jitc-2024-660
47. □ Wellhausen N, Agarwal S, Rommel PC, Gill SI. Better living through chemistry: CRISPR/Cas engineered T cells for cancer immunotherapy. *Curr Opin Immunol.* 2022;74:141-152. doi:10.1016/j.coi.2021.08.006
48. □ Stadtmauer EA, Fraietta JA, Davis MM, Cohen AD. CRISPR-engineered T cells in patients with refractory cancer. *Science.* 2020;367(6481):eaba7365. doi:10.1126/science.aba7365
49. □ Dimitri A, Herbst F, Fraietta JA. Engineering the next-generation of CAR T-cells with CRISPR-Cas9 gene editing. *Mol Cancer.* 2022;21(1):73. doi:10.1186/s12943-022-01559-z
50. Nauman M, Jung Y, Karadal-Ferrena B, Duran C. Requirement of MenaINV for metastatic colonization of the lungs by breast cancer stem cells. *Cancer Res.* 2024;84(22 Suppl):B045. doi:10.1158/1538-7445.AM2024-B045
51. □ Ma Y, Liao J, Cheng H, Yang Q, Yang H. Advanced gene therapy system for the treatment of solid tumours: A review. *Mater Today Bio.* 2024;22:100397. doi:10.1016/j.mtbio.2024.100397
52. □ He P, Yang G, Zhu D, Kong H. Biomolecule-mimetic nanomaterials for photothermal and photodynamic therapy of cancers: Bridging nanobiotechnology and biomedicine. *J Nanobiotechnol.* 2022;20(1):91. doi:10.1186/s12951-022-01691-4
53. □ Wang F, Zhu A, Zhou J, Wang Y, Li J. Near-infrared photoresponsive nanotransducers for precise regulation of gene expression. *Bioconjug Chem.* 2023;34(3):649-662. doi:10.1021/acs.bioconjchem.3c00013
54. Tao J, Bauer DE, Chiarle R. Assessing and advancing the safety of CRISPR-Cas tools: from DNA to RNA editing. *Nat Commun.* 2023;14(1):35886.

55. Koonin EV, Gootenberg JS, Abudayyeh OO. Discovery of diverse CRISPR-Cas systems and expansion of the genome engineering toolbox. *Biochemistry*. 2023;62(1):45-56.
56. Chuang YF, Phipps AJ, Lin FL, et al. Approach for in vivo delivery of CRISPR/Cas system: a recent update and future prospect. *Cell Mol Life Sci*. 2021;78(7):3725.
57. Crowther MD, Svane IM, Met Ö. CRISPR-Cas9 mediated double gene knockout of immune checkpoints PD1 and CISH in tumour-infiltrating T-cells. *J Immunother Cancer*. 2024;12(Suppl 2):A422.
58. D'Amico S, Damiani V, Gragera P, et al. Combining ERAP1 silencing and entinostat therapy to overcome resistance to cancer immunotherapy in neuroblastoma. *J Exp Clin Cancer Res*. 2024;43(1):180.
59. Beavis P. CRISPR/Cas9 engineering of next-generation armoured CAR T cells. *Cancer Immunol Res*. 2024;12(10\_Suppl):PR-02.
60. Zhou Y, Ge Q, Wang X, et al. Advances in lipid nanoparticle-based disease treatment. *ChemMedChem*. 2025;20(5):0938.
61. Fatima H, Singh D, Muhammad H, et al. Improving the use of CRISPR/Cas9 gene editing machinery as a cancer therapeutic tool with the help of nanomedicine. *3 Biotech*. 2025;15(3):04186.
62. Sun J. Nanoparticle therapies: targeted treatment for bladder cancer with reduced side effects. *Int J Nanomedicine*. 2025;20:513952.
63. Nie D, Guo T, Yue M, et al. Research progress on nanoparticles-based CRISPR/Cas9 system for targeted therapy of tumors. *Biomolecules*. 2022;12(9):1239.
64. Lin YQ, Feng KK, Lu JY, et al. CRISPR/Cas9-based application for cancer therapy: Challenges and solutions for non-viral delivery. *J Control Release*. 2023;358:375-389.
65. Yi K, Kong H, Lao YH, et al. Engineered nanomaterials to potentiate CRISPR/Cas9 gene editing for cancer therapy. *Adv Mater*
66. Zhou S, Li Y, Wu Q, Gong C. Nanotechnology-based CRISPR/Cas9 delivery system for genome editing in cancer treatment. *MedCommBiomater Appl*. 2024;5:e70.
67. Singh D. Revolutionizing lung cancer treatment: Innovative CRISPR-Cas9 delivery strategies. *AAPS PharmSciTech*. 2024;25(1):02834.
68. Sims RA, Foley RA, Duggan EC, et al. Delivering the CRISPR/Cas9 system for engineering gene therapies: Recent cargo and delivery approaches for clinical translation. *Front BioengBiotechnol*. 2022;10:973326.
69. Zhang P, Xiao Y, Sun X, et al. Cancer nanomedicine toward clinical translation: Obstacles, opportunities, and future prospects. *Med*. 2023;4(11):817-831.
70. Singh D. Revolutionizing lung cancer treatment: Innovative CRISPR-Cas9 delivery strategies. *AAPS PharmSciTech*. 2024;25(1):02834.
71. Zhou Y, Ge Q, Wang X, et al. Advances in lipid nanoparticle-based disease treatment. *ChemMedChem*. 2025;20(5):0938.
72. Selvarajan V, Gan SKE, Ng YL. Unlocking the potential of cell therapy: exploring cell types, induction methods, and culture techniques. *Front Cell Dev Biol*. 2024;12:1515978.

73. Sabir F, Zeeshan M, Laraib U, Barani M, Rahdar A. DNA-based and stimuli-responsive smart nanocarrier for diagnosis and treatment of cancer: Applications and challenges. *Cancers*. 2021;13(14):3396.
74. Lin M, Qi X. Advances and challenges of stimuli-responsive nucleic acids delivery system in gene therapy. *Pharmaceutics*. 2023;15(5):1450.
75. Deng S, Li X, Liu S, et al. Codelivery of CRISPR-Cas9 and chlorin e6 for spatially controlled tumor-specific gene editing with synergistic drug effects. *Sci Adv*. 2020;6(41):eabb4005.
76. Iqbal Z, Rehman K, Xia J, Shabbir M, Zaman M. Biomaterial-assisted targeted and controlled delivery of CRISPR/Cas9 for precise gene editing. *Biomaterials Sci*. 2023;11(9):2467-2483.
77. Puig Saus C. Longitudinal landscape analysis of the neoantigen-specific T-cell responses in patients with melanoma with or without response to PD1 blockade. *Cancer Immunol Res*. 2025;13(2\_Suppl):IA01.
78. Khan A, Barapatre AR, Babar N, Doshi J. Genomic medicine and personalized treatment: a narrative review. *Ann Med Surg*. 2025.
79. Zhong S, Börgeling Y, Zardo P, et al. Comprehensive transcriptome, miRNA, and kinome profiling identifies new treatment options for personalized lung cancer therapy. *Clin Transl Med*. 2025;15(1):70177.
80. Chen D, Feng X, Li Z, et al. CRISPR/Cas9 technology for advancements in cancer immunotherapy. *Hematol Oncol*. 2024;12(3):00570.
81. Khalil A. Precision oncology in the era of CRISPR-Cas9 technology. *Front Genet*. 2024;15:1506627.
82. Zhang N, He W, Bai M. CRISPR: The game changer in gene and cell therapy. *Front Genet*. 2024;15:1517298.
83. Jalali A, Siddik AB, Yu K, et al. PI3 kinase and checkpoint inhibition: sexual dimorphism in a native model of glioblastoma (GBM). *J Immunother Cancer*. 2024;12(Suppl 2):A872.
84. Wang M, Jia L, Dai X, Zhang X. Advanced strategies in improving the immunotherapeutic effect of CAR-T cell therapy. *Mol Oncol*. 2024.
85. Chen D, Feng X, Li Z, et al. CRISPR/Cas9 technology for advancements in cancer immunotherapy. *Hematol Oncol*. 2024.
86. Xuan Y, Yan W, Wang R, et al. GM-CSF and IL-21-armed oncolytic vaccinia virus significantly enhances anti-tumor activity and synergizes with anti-PD1 immunotherapy in pancreatic cancer. *Front Immunol*. 2025.
87. Youssef E, Fletcher B, Palmer D. Enhancing precision in cancer treatment: the role of gene therapy and immune modulation in oncology. *Front Med*. 2025
88. Gameel FM. Exploring the Dual Roles of RNAs as Oncogenic Drivers and Therapeutic Targets in Cancer: A Cross-Sectional Study. *Mol Cancer Ther*. 2024;23(11\_Suppl):B008.
89. Mukharya A, Nikam AN, Roy AA, Pokale R. Synergizing CRISPR-Cas9 with Advanced Artificial Intelligence and Machine Learning for Precision Drug Delivery: Technological Nexus and Regulatory Insights. *PubMed*. 2024.

90. Wu H. Advances in the Application of CRISPR-Cas9 in Stem Cell Therapy. *Theor Nat Sci.* 2025.
91. Hwang WYK, Muzammil EM. The Current State of Cytotherapy and the Field of Cell and Gene Therapy. *Cytotherapy.* 2025.
92. IKE PC, KELECHI OA, AGBOLI VI. The Future of Medicine: Advancing Gene Therapy with CRISPR-Cas9's Exact Precision in Pediatric Males. *Systems Research.* 2025.
93. He W, Bai M. CRISPR: The Game Changer in Gene and Cell Therapy. *Front Genet.* 2024.
94. Li R, Yang F, Chu B, et al. Exploring Retinal Degenerative Diseases through CRISPR-Based Screening. *Mol Biol Rep.* 2024
95. Haq EU, Yousaf A, Sardar M, Irshad A. Uncovering Genetic Interactions: CRISPR-Mediated Gene Knockouts and Activations in Understanding Complex Diseases. *Pak-Euro J Med Life Sci.* 2024.
96. Khalil A. Precision oncology in the era of CRISPR-Cas9 technology. *Front Genet.* 2024.
97. Wang D, Xing Q. The Roles and Features of Genetic Variants in Children with Cerebral Palsy. *J Bio-X Res.* 2024.
98. Vickram AS, Manikandan S, Richard T, et al. Targeted Gene Therapy: Promises and Challenges in Disease Management. *J Bio-X Res.* 2024.
99. Mamadapur SA, Jadhav HY. Innovations in skin cancer treatment: The role of nanotechnology and cellular biotechnology. *J Cell Biotechnol.* 2025.
100. Zou C, Liu X, Wang W, et al. Targeting GDF15 to enhance immunotherapy efficacy in glioblastoma through tumor microenvironment-responsive CRISPR-Cas9 nanoparticles. *J Exp Clin Cancer Res.* 2025.
101. Fatima H, Singh D, Muhammad H, et al. Improving the use of CRISPR/Cas9 gene editing machinery as a cancer therapeutic tool with the help of nanomedicine. *3 Biotech.* 2025.
102. Sun J. Nanoparticle therapies: targeted treatment for bladder cancer with reduced side effects. *Int J Nanomedicine.* 2025.
103. Zhou Y, Ge Q, Wang X, et al. Advances in lipid nanoparticle-based disease treatment. *ChemMedChem.* 2025;20(5):0938.
104. Yi K, Kong H, Lao YH, et al. Engineered nanomaterials to potentiate CRISPR/Cas9 gene editing for cancer therapy. *Adv Mater.* 2024;36(11):2300665.
105. Saadh MJ, Khidr WA, Alfarttoosi KH, Bishoyi AK. Metal nanoparticles as a promising therapeutic approach for prostate cancer diagnosis and therapy: a comprehensive review. *Med Oncol.* 2025.
106. Kumar N, Ranjan OP. Emerging nanocarriers as advanced delivery tools for the treatment of leukemia. *Nanomedicine.* 2025.
107. Elumalai K, Srinivasan S. Harnessing Nanoparticle Technology for Precision Medicine in Head and Neck Cancer: Targeted Delivery, Immunomodulation, and Clinical Translation. *Nano TransMed.* 2025.
108. Kang H, Xu W, Guan G, et al. Chemical Design of Magnetic Nanomaterials for Imaging and Ferroptosis-Based Cancer Therapy. *Chem Rev.* 2025.

109. Singh D. Revolutionizing Lung Cancer Treatment: Innovative CRISPR-Cas9 Delivery Strategies. *AAPS PharmSciTech*. 2024.
  110. Fatima H, Singh D, Muhammad H, et al. Improving the use of CRISPR/Cas9 gene editing machinery as a cancer therapeutic tool with the help of nanomedicine. *3 Biotech*. 2025.
  111. Dubey AK, Mostafavi E. Biomaterials-mediated CRISPR/Cas9 delivery: Recent challenges and opportunities in gene therapy. *Front Chem*. 2023.
  112. Zhang P, Xiao Y, Sun X, et al. Cancer nanomedicine toward clinical translation: Obstacles, opportunities, and future prospects. *Med*. 2023;4(11):817-831.
  113. Eskandar K. Nanotechnology in Cancer Treatment: Innovative Approaches to Overcoming Drug Resistance in Tumors. *Indonesian Journal of Cancer Chemoprevention*. 2025.
- 

---

Received: May 15<sup>th</sup> 2025,

Accepted: May 28<sup>th</sup> 2025

Licensee Abhipublications *Open*.

This is an open access article licensed under the terms of the Creative Commons Attribution Non- Commercial License (<http://www.abhipublications.org/ijpe>) which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited

---